

A new statistical approach for the simultaneous clustering of genes and cells in spatial transcriptomic experiments

Andrea Sottosanti

Davide Risso

December 14th, 2020

University of Padova - Department of Statistical Sciences

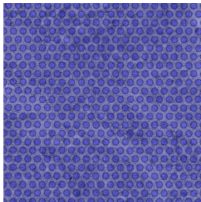
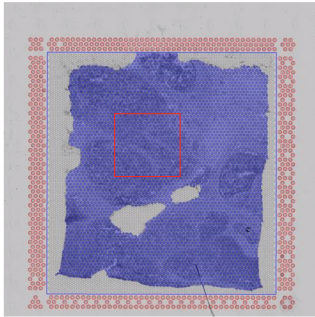
andrea.sottosanti@unipd.it



UNIVERSITÀ
DEGLI STUDI
DI PADOVA



The 10x Genomics Visium technology



3,813

Number of Spots Under Tissue

149,800

Mean Reads per Spot

5,394

Median Genes per Spot

- $j = 1, \dots, p$ spots, each of which is spatially located;
- Number of spots \approx number of cells;
- for each spot, $i = 1, \dots, n$ gene expressions are available.

Spatially expressed genes and research motivations

- The rise of such advanced technology has increased the interest for the so-called *spatially expressed* (*s.e.*) genes.

Spatially expressed genes and research motivations

- The rise of such advanced technology has increased the interest for the so-called *spatially expressed* (*s.e.*) genes.
- There are methods for discovering *s.e.* genes: spatialDE [Svensson et al., 2018], Trendsceek [Edsgård et al., 2018], SPARK [Sun et al., 2020].

Spatially expressed genes and research motivations

- The rise of such advanced technology has increased the interest for the so-called *spatially expressed (s.e.) genes*.
- There are methods for discovering s.e. genes: spatialDE [Svensson et al., 2018], Trendsceek [Edsgård et al., 2018], SPARK [Sun et al., 2020].

however...

- These methods do not account for the presence of different cell types.
- Some (clusters of) genes might be s.e. just in some specific cell types.

Some aspects to consider

- Let

$\mathbf{X}_{n \times p}$: x_{ij} = measure of expression of the i -th gene
in the j -th spot.

Some aspects to consider

- Let

$\mathbf{X}_{n \times p}$: x_{ij} = measure of expression of the i -th gene
in the j -th spot.

- The spatial coordinates of the spots (s_1, \dots, s_p) are known.

Some aspects to consider

- Let

$\mathbf{X}_{n \times p}$: x_{ij} = measure of expression of the i -th gene
in the j -th spot.

- The spatial coordinates of the spots (s_1, \dots, s_p) are known.
- Just for now, we assume there is only one type of cells.

Some aspects to consider

- Let

$\mathbf{X}_{n \times p}$: x_{ij} = measure of expression of the i -th gene
in the j -th spot.

- The spatial coordinates of the spots (s_1, \dots, s_p) are known.
- Just for now, we assume there is only one type of cells.

Some aspects to consider:

1. correlation of the genes $\rightarrow \text{Cor}(x_{i\textcolor{red}{j}}, x_{i'\textcolor{red}{j}}),$
2. (spatial) correlation of the spots $\rightarrow \text{Cor}(x_{i\textcolor{red}{j}}, x_{i\textcolor{red}{j'}}).$

A statistical model

We assume that the experiment matrix \mathbf{X} distributes as

$$\mathbf{X} \sim \mathcal{MVN}_{n,p}(\boldsymbol{\mu}, \boldsymbol{\Sigma}, \boldsymbol{\Delta}),$$

where \mathcal{MVN} denotes the [Matrix Variate Normal](#) distribution [Gupta and Nagar, 2018]:

- $\boldsymbol{\mu} = \mu \cdot \mathbf{1}_{n \times p}$ is the mean matrix;
- $\boldsymbol{\Sigma}$ is an $n \times n$ matrix which express the **correlation of the genes** (rows);
- $\boldsymbol{\Delta}$ is an $p \times p$ matrix which express the **correlation of the cells** (columns).

A statistical model

Regarding the rows,

$$\Sigma := \begin{cases} \sigma_i^2 & \text{in position } (i, i); \\ 0 & \text{elsewhere;} \end{cases}$$

$$\sigma_i^2 \sim \mathcal{IG}(\alpha, \beta).$$

A statistical model

Regarding the rows,

$$\Sigma := \begin{cases} \sigma_i^2 & \text{in position } (i, i); \\ 0 & \text{elsewhere;} \end{cases} \quad \sigma_i^2 \sim \mathcal{IG}(\alpha, \beta).$$

Regarding the columns,

$$\Delta = \tau \cdot \mathbf{K}(\phi) + \xi \cdot \mathbf{1}_{p \times p}.$$

- $\tau \in \mathbb{R}^+$ is the amount of spatial expression;
- $\mathbf{K}(\cdot)$ is the spatial kernel matrix: example,

$$\mathbf{K}_{j,j'} = \exp\{-\|\mathbf{s}_j - \mathbf{s}_{j'}\|^2 / (2\phi^2)\};$$

- $\phi \in \mathbb{R}^+$ is the spatial scale;
- $\xi \in \mathbb{R}^+$ is the nugget effect (variance not imputable to the spatial structure);

The Co-clustering problem

- K gene clusters $\rightarrow \mathcal{C}_i = k$ means that gene i belongs to the k -th gene cluster;
- R cell clusters $\rightarrow \mathcal{D}_j = r$ means that cell j belongs to the r -th cell type.

The Co-clustering problem

- K gene clusters $\rightarrow \mathcal{C}_i = k$ means that gene i belongs to the k -th gene cluster;
- R cell clusters $\rightarrow \mathcal{D}_j = r$ means that cell j belongs to the r -th cell type.

	$r = 1$	$r = 2$	\dots	$r = R$
$k = 1$	\mathbf{X}_{11}	\mathbf{X}_{12}	\dots	\mathbf{X}_{1R}
$k = 2$	\mathbf{X}_{21}	\ddots	\dots	\vdots
\dots	\vdots	\dots	\ddots	\vdots
$k = K$	\mathbf{X}_{K1}	\dots	\dots	\mathbf{X}_{KR}

The Co-clustering problem

- K gene clusters $\rightarrow \mathcal{C}_i = k$ means that gene i belongs to the k -th gene cluster;
- R cell clusters $\rightarrow \mathcal{D}_j = r$ means that cell j belongs to the r -th cell type.

$$\mathbf{X} = \begin{array}{c} \begin{array}{ccccc} & r = 1 & r = 2 & \dots & r = R \end{array} \\ \begin{array}{c} k = 1 \\ k = 2 \\ \dots \\ k = K \end{array} \end{array} \begin{array}{|c|c|c|c|} \hline \mathbf{X}_{11} & \mathbf{X}_{12} & \dots & \mathbf{X}_{1R} \\ \hline \mathbf{X}_{21} & \ddots & \dots & \vdots \\ \hline \vdots & \dots & \ddots & \vdots \\ \hline \mathbf{X}_{K1} & \dots & \dots & \mathbf{X}_{KR} \\ \hline \end{array}$$

\Downarrow

$$\mathbf{X}_{kr} \sim \mathcal{MVN}_{n_k, p_r}(\boldsymbol{\mu}_{kr}, \boldsymbol{\Sigma}_{kr}, \tau_{kr} \cdot \mathbf{K}(\phi_r) + \xi_{kr} \cdot \mathbb{1}_{p_r \times p_r}),$$

$$\sigma_{kr,i}^2 \sim \mathcal{IG}(\alpha_{kr}, \beta_{kr})$$

for $k = 1, \dots, K$ and $r = 1, \dots, R$.

- We exploit the human dorsolateral prefrontal cortex (DLPFC) spatial transcriptomics data generated with the 10x Genomics Visium technology by [Maynard et al., 2020] and contained in the R package spatialLIBD [Collado-Torres et al., 2020].
- We reduced the dataset size, using the first **1000** most variable **genes** measured in **1585 spots**.
- We run our model on log-counts data using $K = 1$ and $R = 4$.
- The estimation procedure is initialized using the results from k-means.

spatialLIBD data - clustering

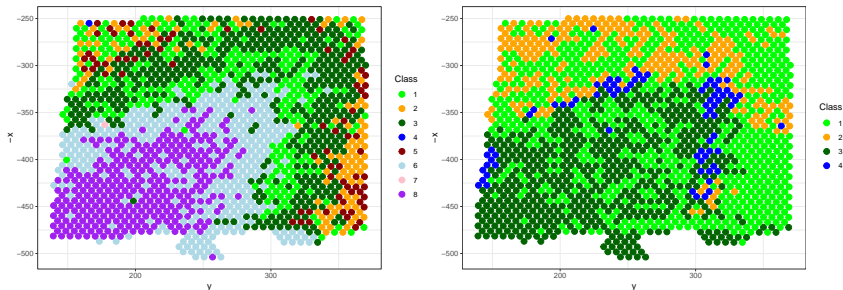


Figure 1: Data: subject 151673. Left: clustering provided by spatialLIBD. Right: Clustering from our method.

cell cluster	$\hat{\mu}$	$\hat{\tau}/\hat{\xi}$	$\hat{\phi}$
1	0.863	0.479	19.159
2	0.451	0.304	21.232
3	0.501	0.357	19.283
4	0.198	0.200	31.440

spatialLIBD data - genes variance

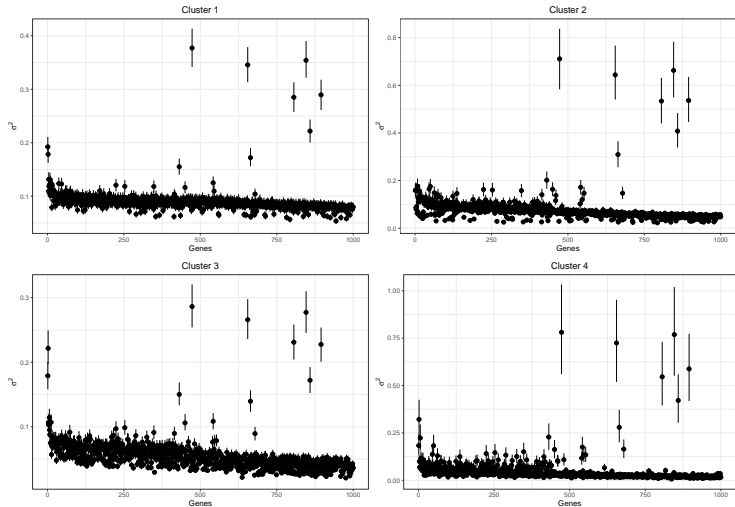


Figure 2: Expected value and 95% interval of σ_i^2 in every cell cluster, given the data the parameter estimates. The first two highly variable genes are ENSG00000123560 and ENSG00000197971.



- Dario Righelli
- Martin Morgan, Vince Carey, Levi Waldron
- Giovanna Menardi

Thank you for the attention!

andrea.sottosanti@unipd.it



Collado-Torres, L., Maynard, K. R., and Jaffe, A. E. (2020).
LIBD Visium spatial transcriptomics human pilot data inspector.

<https://github.com/LieberInstitute/spatialLIBD> - R package
version 1.2.0.



Edsgård, D., Johnsson, P., and Sandberg, R. (2018).
**Identification of spatial expression trends in single-cell
gene expression data.**

Nature methods, 15(5):339–342.

References II



Gupta, A. K. and Nagar, D. K. (2018).

Matrix variate distributions, volume 104.

CRC Press.



Maynard, K. E., Collado-Torres, L., Weber, L. M., Uytingco, C., Barry, B. K., Williams, S. R., et al. (2020).

Transcriptome-scale spatial gene expression in the human dorsolateral prefrontal cortex.

bioRxiv.



Sun, S., Zhu, J., and Zhou, X. (2020).

Statistical analysis of spatial expression patterns for spatially resolved transcriptomic studies.

Nature Methods, 17(2):193–200.



Svensson, V., Teichmann, S. A., and Stegle, O. (2018).

SpatialDE: identification of spatially variable genes.

Nature methods, 15(5):343–346.